Kaiyi Jiang

kaiyi@mit.edu | Cambridge, MA, 02139 | 865-307-4066

EDUCATION

Massachusetts Institute of Technology, Cambridge, MA

Expected May 2025

Doctor of Philosophy (Ph.D.) in Biological Engineering

Thesis title: Harnessing biological diversity and machine learning to develop a cell programming toolbox

Advisors: Omar Abudayyeh, Jonathan Gootenberg, and Michael Birnbaum

Rice University, Houston, TX

Bachelor of Engineering in Biomedical Engineering (Summa cum laude)

May 2021

Thesis title: Engineering synthetic phosphorylation signaling networks in mammalian cells

Advisors: Caleb Bashor, Pankaj Mehta, and Gang Bao

RESEARCH INTERESTS

- Harnessing biological diversity to discover novel reprogrammable systems
- AI/ML models to engineer RNA and proteins
- Synthetic biology for in vivo recording and lineage tracing
- Gene/cell therapy for autoimmune disorders, aging, and cancer

PUBLICATIONS & PREPRINTS (* denotes co-first author)

- [1] <u>Jiang, K.*</u>, Yan, Z.*, Bernardo, MD., Sgrizzi, S.R., Villiger, L., Kayabolen, A., Kim, B., Carscadden, J.K., Hiraizumi, M., Nishimasu, H., Gootebnerg, J.S., Abudayyeh, O.O. (2024). Rapid protein evolution by few-shot learning with a protein language model. *bioRxiv*. [In Revision]
 - o Summary:
 - I developed EVOLVE-Pro, a state-of-the-art (SOTA) ensemble protein language model that can rapidly design protein variants with up to 100-fold higher activity than wild-type. We showed EVOLVE-Pro's SOTA performance across 12 DMS datasets. We used it to evolve a T7 RNA polymerase that is up to 100-fold better than wild-type enzymes and demonstrated efficient in vivo gene therapy and mRNA therapeutics with these engineered enzymes.
- [2] Koob, J.*, **Jiang, K.***, Sgrizzi, S.R.*, Chen, F., Abudayyeh, O.O., Gootenberg, J.S. (2024). Sensing and perturbing mammalian cell states with reprogrammable ADAR sensors. [AIP at *Nature Protocols*]
- [3] Yang, X., Rocks, JW., <u>Jiang, K.</u>, Walters, AJ., Rai, K., Liu, J., Nguyen, J., Olson, SD., Mehta, P., Collins, JJ., Daringer, NM., Bashor, CJ. (2024). Engineering synthetic phosphorylation signaling networks in human cells. [AIP at *Science*]
- [4] <u>Jiang, K.*</u>, Koob, J.*, Chen, X.D.*, Krajeski, R.N.*, Zhang, Y., Volf, V., Zhou, W., Sgrizzi, S.R., Villiger, L., Gootenberg, J.S., Chen, F., Abudayyeh, O.O. (2023). Programmable eukaryotic protein synthesis with RNA sensors by harnessing ADAR. *Nature Biotechnology*.
 - o Summary:
 - We engineered the first robust mammalian RNA sensor based on ADAR called RADARS. The sensor can be reprogrammed to track any RNA species inside eukaryotic cells and allow conditional cargo expression based on the presence/expression of target mRNA(s). We demonstrate that the system can be readily integrated into AAV, lentivirus, and synthetic mRNA to selectively turn on an arbitrary protein of interest. We showcase the use of this system in cell-specific killing, lineage tracing, and *in vivo* recording for reprogrammable cell control.
 - Highlight:
 - This work was highlighted in Derek Lowe's IN THE PIPELINE. (2022), Ono et al, RNA Biology. (2023) and Twist Bioscience's Top Moments in Biotech (2022)
- [5] <u>Jiang, K.*</u>, Lim, J.*, Sgrizzi, S.R., Trinh, M., Kayabolen, A., Yutin, N., Bao, W., Kato, K., Koonin, E., Gootenberg, J.S., Abudayyeh, O.O. (2023). Programmable RNA-guided DNA endonucleases are widespread in eukaryotes. *Science Advances*.
 - o Summary:
 - We discovered a novel group of RNA-guided DNA endonucleases widespread in eukaryotes and their viruses named Fanzor. This was the first evidence of reprogrammable RNA-guided DNA nucleases in eukaryotes, and we characterized Fanzors' (eukaryotic homologs of TnpB) adaptations into the eukaryotic world with nuclear localization signal (NLS), introns and association with diverse transposons. This work established the presence of RNA-guided DNA nucleases in all three kingdoms of life and the diversity of RNA-guided reprogrammable systems.
 - Highlight:
 - This work was highlighted in Patinios et al, Mol. Cell. (2023) & Karvelis et al, The CRISPR Journal. (2023)
- [6] Kato, K.*, Okazaki, S.*, Schmitt-Ulms, C.*, <u>Jiang, K.*</u>, Zhou, W., Ishikawa, J., Isayama, Y., Adachi, S., Nishizawa, T., Makarova, K.S., et al. (2022). RNA-triggered protein cleavage and cell death by the RNA-guided type III-E CRISPR-Cas nuclease-protease complex. *Science*.
 - o Summary:
 - We discovered the first RNA-guided protease systems in prokaryotic antiviral defense systems. We biochemically characterized the Cas7-11/Csx29/Csx30 systems in the context of an abortive infection module against phage invasion. We then engineered the system and adapted it as an RNA-sensor system in mammalian cells.
 - Highlight:
 - This work was highlighted in Burgess et al, Nature Reviews Genetics. (2023), Chen et al, Trends in microbiology. (2023), and Wang et al, Nucleic Acids Research. (2022).

- [7] Yarnall, M.T.N.*, Ioannidi, E.I.*, Schmitt-Ulms, C.*, Krajeski, R.N.*, Lim, J., Villiger, L., Zhou, W., Jiang, K., Roberts, N., Zhang, L., et al. (2022). Drag-and-drop genome insertion without DNA cleavage with CRISPR-directed integrases. *Nature* Biotechnology.
- [8] Sebesta, C., Torres Hinojosa, D., Wang, B., Asfouri, J., Li, Z., Duret, G., Jiang, K., Xiao, Z., Zhang, L., Zhang, Q., et al. (2022). Subsecond multichannel magnetic control of select neural circuits in freely moving flies. Nature Materials.
- [9] Zhang, L., Zhang, Q., Hinojosa, D.T., Jiang, K., Pham, Q.K., Xiao, Z., Colvin, V.L., Bao, G. (2022) Multifunctional Magnetic Nanoclusters Can Induce Immunogenic Cell Death and Suppress Tumor Recurrence and Metastasis. ACS Nano.
- [10] Jiang, K., Zhang, Q., Hinojosa, D.T., Zhang, L., Xiao, Z., Yin, Y., Tong, S., Colvin, V.L., Bao, G. (2021) Controlled oxidation and surface modification increase heating capacity of magnetic iron oxide nanoparticles. Applied Physics Reviews.
- [11] Jiang, K., Zhang, L., Bao, G. (2021) Magnetic iron oxide nanoparticles for biomedical applications. *Current Opinion in* Biomedical Engineering.

INVITED TALKS & PRESENTATIONS

- 2024 ML Protein Engineering Seminar Series. (Virtual)
- 2024 Cradle Bio invited seminar on machine learning for protein engineering. (Virtual)
- 2024 Harvard Medical School Machine learning to accelerate biology journal club, Boston, MA.
- 2024 SynBYSS, Rising Star junior speaker. (Virtual)
- 2024 Bunker Hill community college, STEM Seminar, Boston, MA.
- 2024 Tufts BME162 Molecular Biotech invited lecture, Boston, MA.
- 2023 Bioengineering and Toxicology Seminar, Boston, MA.
- 2023 Harvard Medical School Genome Engineering Symposium, Boston, MA.
- 2023 Broad Gene regulation observatory seminar, Boston, MA.
- 2023 Mammalian Synthetic Biology Workshop (Poster presentation), Stanford, CA.
- 2023 Broad Institute Cell Circuits and Epigenetics Seminar, Boston, MA.
- 2022 Harvard Medical School Genome Engineering Seminar Series, Boston, MA. (Virtual)
- 2022 Boston Mammalian Synthetic Biology Symposium, Boston, MA.
- 2022 Single Cell Genomics Day, NYC, NY. (Virtual)
- 2019 Annual Meeting of Biomedical Engineering Society (BMES), Philadelphia, PA.

PROFESSIONAL EXPERIENCE	
Graduate Student, Abudayyeh-Gootenberg Lab, MIT & Harvard Medical School	2021-2025
 Developed a robust cell state sensor RADARS 	
• Characterized the first RNA-guided protease (Craspase) and eukaryotic RNA-guided nuclease (Fanzor)	
 Developed a robust few-shot protein evolution language model (EVOLVEpro) 	
Rotational student with Prof. James Collins, MIT	2021
Rotational student with Prof. Michael Birnbaum, MIT	2021
Undergraduate Research Assistant, Caleb Bashor Lab, Rice University	2018-2021
 Engineered a fully synthetic phosphorylation network in mammalian cell 	
Intern, Regeneron Pharmaceuticals (Therapeutic antibody group)	2019
 Elucidated molecular mechanisms driving different humoral responses for adjuvants 	
Undergraduate Research Assistant, Gang Bao Lab, Rice University	2017-2021
 Designed a nanoparticle system for in vivo hyperthermia therapy 	

AWARDS & HONORS

11 () 11 11 12 D GG 11 O1 (O1 D	
• Innovators Under 35 (TR35), MIT Technology Review, Asia-Pacific Region	2024
• 30 under 30, All-America Chinese Youth Federation, The Log Angeles Post	2024
The Wishnok Prize, Massachusetts Institute of Technology	2024
MiraclePlus (Former Y Combinator China) Fellow	2024
Distinction in Research and Creative Works	2021
Outstanding Junior in Bioengineering	2020
Tau Beta Pi Member	2019
• Louis J. Walsh Scholarships	2019-2021
Best Oral Presentation Award, Rice University Research Symposium	2019
President's Honor Roll	2017-2021

PATENTS

- Abudayyeh, O., Gootenberg, J., Nishimasu, H., Kazuki, K., Okazaki, S., Schmitt-Ulms, C., and Jiang, K. (2024). 1. RNA-triggered protein cleavage and applications by the CRISPR Cas7-11-Csx29 complex. US Patent App. 18/234,690
- Jiang, K., Krajeski, R.N., Abudayyeh, O.O., Gootenberg, J.S., Zhang, Y., Chen, F., Chen, X., and Koob, J.G. 2. (2023). Deaminase-based rna sensors. US Patent App. 17/806,879
- 3. Abudayyeh, O., Gootenberg, J., Villiger, L., and Jiang, K. (2023). Programmable insertion approaches via reverse transcriptase recruitment. US Patent App. 18/067,214

- 4. Abudayyeh, O., Gootenberg, J., and **Jiang, K**. (2023). Site specific genetic engineering utilizing trans-template rnas. US Patent App. 18/303,533
- 5. **Jiang, K.,** Villiger, L., Abudayyeh, O., and Gootenberg, J. (2023) Systems, methods, and compositions comprising miniature CRISPR nucleases for gene editing and programmable gene activation and inhibition. US Patent App. 63/211.610
- 6. Abudayyeh, O., Gootenberg, J., **Jiang, K**, and Lim, J. (2023). Fanzors are rna-guided nucleases encoded in eukaryotic genomes. US Patent App. 18/406,066

TEACHING EXPERIENCE

•	Fall 2022	20.110J: Thermodynamics of Biomolecular Systems, Massachusetts Institute of Technology
		Graduate Teaching Assistant for Prof. Christopher Voigt, Prof. Linda Griffith & Prof. Eric Alm
•	Fall 2020	BIOE252: Bioengineering Fundamentals, Rice University
		Undergraduate Teaching Assistant for Prof. Renata Ramos
•	Fall 2019	MATH211: ODEs and Linear Algebra, Rice University
		Undergraduate Teaching Assistant for Prof. Milivoje Lukic

MENTORSHIP

• For research associate Josephine Carscadden

May 2024-Now

Josephine worked with me on evolving antibodies with machine learning models.

• For high school intern Katrina Yang

May 2024-Aug 2024

Katrina worked with me on optimizing circular RNA production.

• For undergraduate intern Rahul Rajendran

May 2024-Aug 2024

Rahul worked with me on optimizing large cargo gene insertion technology

• For rotational student Jett Liu (MIT Microbiology)

Jan-March2024

Jett worked with me on using a large protein language model to optimize Rubisco's catalytic activity.

• For rotational student Kai Wang (Harvard BBS)

Jan- March2024

Kai worked with me on exploring RNA gated expression of cytokines and optimized the production of RADARS synthetic mRNA for in vivo delivery.

• For research associate Samantha Sgrizzi

2022-2024

Sam worked with me on using machine learning models to in silico evolve higher activity proteins including polymerases, CRISPR/Cas nucleases, and carbonic anhydrases. She developed high throughput cell free expression systems for testing mutants. Sam is now a PhD student in University of Washington, Seattle.

• For undergraduate student Michael Trinh (University of Toronto)

Jan-Dec2023

Michael worked with me discovery of eukaryotic RNA-guided nucleases. He also designed RADARS constructs that selectively turn on OSKM in senescent cells. With this work, Michael won the time initiative aging research fellowship. Michael is now a PhD student in McGill University.

For high school student Rayya Reda Frayn

June-August 2023

Rayya is learning basic molecular biology skills and looking into cost-effective high throughput multi-part Gibson assembly.

• For rotational student Benyapa Khowpinitchai (Harvard BBS)

Feb-June2023

Ben worked with me on engineering miniature CRISPR/Cas12f nucleases for *in vivo* gene regulation.

• For rotational student Shuchen Luo (MIT Chemistry)

Aug- Dec2022

Shuchen worked with me on optimization of RADARS platform for cancer cell specific killing and detection.

• For rotational student Kathrin Kajderowicz (MIT BCS)

Jan- March2024

Kat worked with me on using RADARS to perform RNA gated genome editing that can be used for *in vivo* lineage tracing. Kat is now a PD Soros Ph.D. Fellow at Whitehead Institute.

Professional Societies

Biomedical Engineering Society (BMES)

2020-Now 2023-Now

• American Institute of Chemical Engineers (AIChE)

Journal Reviewer

*PLOS ONE *Frontiers in Neuroscience *Science of the Total Environment